

SWITCHING ANDROGEN RECEPTOR ANTAGONISTS TO AGONISTS BY MODIFYING C-RING SUBSTITUENTS ON PIPERIDINO[3,2-*g*]QUINOLINONE

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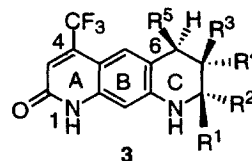
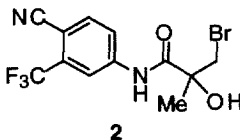
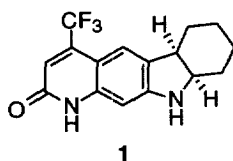
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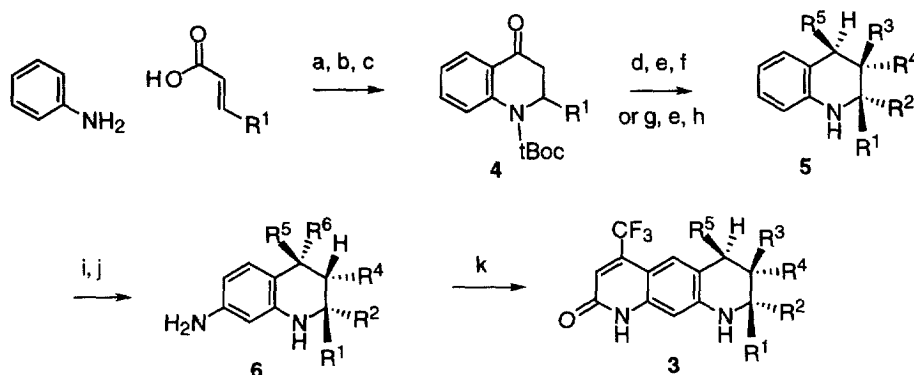
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Abstract: New nonsteroidal human androgen receptor (hAR) agonists were developed from an hAR antagonist pharmacophore, 2(1*H*)-piperidino[3,2-*g*]quinolinone. (±)-*trans*-7,8-Diethyl-4-trifluoromethyl-2(1*H*)-piperidino[3,2-*g*]quinolinone was synthesized and demonstrated potent hAR agonist activity ($EC_{50} = 3$ nM) in the cell-based cotransfection assay and high binding affinity ($K_i = 16$ nM) in the competitive receptor binding assay. © 1999 Elsevier Science Ltd. All rights reserved.

Introduction: In contrast to nonsteroidal antiandrogens, which have been successfully used in clinic,² nonsteroidal androgens had not been known in literature until recently. Both of the reported new nonsteroidal androgens (1 and 2)³ were derived from analoguing antiandrogen pharmacophores. In our effort to develop novel human androgen receptor (hAR) modulators we reported the discovery of a linear tricyclic hAR antagonist pharmacophore, 2(1*H*)-piperidino[3,2-*g*]quinolinone (3).⁴ To take advantage of this orally available pharmacophore, we systematically investigated the substitution pattern at the C-ring.



Chemistry: The analogues of generic structure 3 ($R^2 = H$) were prepared by the synthetic sequence shown in Scheme 1. The preparation of 4 started from conjugate addition of aniline to an olefinic acid and a PPA mediated cyclization followed by nitrogen protection with a *t*-Boc group. The 2,4-disubstituted tetrahydroquinolines (5, $R^2 = R^3 = R^4 = H$) were synthesized by Grignard addition to 4 and deprotection of *t*-Boc followed by palladium catalyzed hydrogenation in the presence of an acid. The 2,3-di- or 2,3,3-trisubstituted tetrahydroquinolines (5, $R^2 = R^5 = H$) were prepared by alkylation of 4 with iodoalkane and deprotection of *t*-Boc followed by BF_3 catalyzed Et_3SiH reduction. Nitration of 5 followed by palladium catalyzed hydrogenation provided aminoquinolines 6, which were subsequently converted to 3 via Knorr cyclization.⁵ Stereoisomers were separated by HPLC and chiral compounds were tested as racemates. The preparation of compounds 24 and 25 are illustrative.⁶

Scheme 1^a

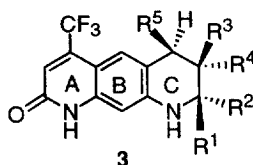
^aReagents: (a) toluene, reflux, 20 h; (b) PPA, 110 °C, 6 h; (c) *t*-Boc₂O, DMAP, THF, rt, 6 h; (d) R⁵MgX, THF, rt; (e) TFA, CH₂Cl₂, rt, 30 min; (f) Pd/C, H₂, EtOAc, H₂SO₄, rt, 15 h; (g) R³I or R⁴I, NaH, DMF, rt, 15 h; (h) Et₃SiH, BF₃·OEt₂, CH₂Cl₂, sealed tube, 100 °C, 24 h; (i) H₂SO₄, HNO₃, -10 °C, 10 min; (j) Pd/C, H₂, EtOAc, rt, 15 h; (k) EtO₂CCH₂COCF₃, ZnCl₂, EtOH, reflux, 8 h.

Results and Discussion: A number of 8,8-dialkyl analogues have shown potent hAR antagonist activity in the cell-based cotransfection assay and high affinity in the receptor competitive binding assay, in addition, compound 7 demonstrated potent *in vivo* antiandrogenic activity following oral administration.⁴ To reveal additional structure–activity relationship (SAR) of the pharmacophore, we continued to use the cotransfection and the binding assays as guides. Our initial attempt of removing one of the 8-alkyl substituents resulted in four analogues. Compounds 8 and 9 showed antagonist efficacy weaker than parent compound 7 but with stronger binding affinity to hAR (see Table 1). Additionally, compound 8 demonstrated potent partial agonist activity. To introduce a small alkyl group at C6, four *cis*-isomers (12–15) of 6,8-dialkyl analogues were prepared by the current synthetic route and all of them gave potent partial agonist activity similar to 8. Interestingly, among the 7,8-dialkyl analogues compounds 22 (LG121091) and 24 (LG121104) showed potent full agonist activity similar to that of dihydrotestosterone in the hAR cotransfection assay. It was noticed that the proper size of 7-alkyl substituent was essential for the full agonist activity (i.e., 7-ethyl or propyl gave agonists and 7-methyl gave partial or full antagonists) and the size of 8-substitution is less crucial. The *trans*-isomers (22 and 24) afforded better agonist activity than their *cis*-isomers (23 and 25) although both have similar binding affinity. The 7,7,8-trialkyl analogues (18, 21 and 26) were also examined and all of them are hAR antagonists.

The cross-reactivity of the new AR agonists 22 and 24 was evaluated using human progesterone, estrogen, glucocorticoid, and mineralocorticoid receptor (hPR, hER, hGR, and hMR) cotransfection assays and only hPR antagonist activity was observed. The IC₅₀ values of 22 and 24 in hPR cotransfection assay are 280 and 260 nM with 78% and 73% efficacy.⁷

Conclusion: Several selective and potent hAR agonists were obtained from a known antagonist pharmacophore **3** (R^1, R^2 = alkyl) by modifying the substituents on the C-ring.⁸ The size and relative stereochemistry at C7/C8 are essential for the agonist activity. This new discovery combined with other results^{3a,9} from our laboratory provided a solid foundation of developing new nonsteroidal hAR agonists for clinical uses.

Table 1. Cotransfection and Competitive Binding Data for the Quinolinone Analogues.^a



#	R ¹	R ²	R ³	R ⁴	R ⁵	hAR Agonist ^b mean ± SEM		hAR Antagonist ^b mean ± SEM		hAR Binding ^c mean ± SEM
						Eff(%)	EC ₅₀ (nM) ^c	Eff(%)	IC ₅₀ (nM) ^c	K _i (nM)
		Dihydrotestosterone				100 ± 0	6 ± 1	- ^d	-	2 ± 0.4
		2-Hydroxyflutamide				-	-	83 ± 1	15 ± 2	27 ± 8
7	Me	Me	H	H	H	-	-	74 ± 2	27 ± 5	26 ± 5
8	Me	H	H	H	H	33 ± 5	21 ± 9	37 ± 8	19 ± 13	7 ± 1
9	Et	H	H	H	H	-	-	55 ± 5	11 ± 2	8 ± 2
10	Pr	H	H	H	H	-	-	84 ± 4	32 ± 6	24 ± 5*
11	i-Pr	H	H	H	H	-	-	82 ± 4	25 ± 9	109 ± 44
12	Me	H	H	H	Me	24 ± 2*	59 ± 9*	52 ± 6	15 ± 6	6 ± 3
13	Me	H	H	H	Et	30 ± 8*	66 ± 35*	-	-	23
14	Et	H	H	H	Et	47 ± 9	32 ± 8	-	-	9 ± 1*
15	Et	H	H	H	Me	37 ± 10	119 ± 72	40 ± 2	137 ± 98	21 ± 8
16	Me	H	H	Me	H	-	-	66 ± 4	33 ± 15	22 ± 6
17	Me	H	Me	H	H	26 ± 4	34 ± 8	41 ± 9	74 ± 48	15 ± 3
18	Me	H	Me	Me	H	-	-	78 ± 5	24 ± 7	82 ± 18
19	Et	H	H	Me	H	31	1668	40 ± 8	11 ± 3	12 ± 0
20	Et	H	Me	H	H	45	2468	66 ± 9	15 ± 6	11 ± 2
21	Et	H	Me	Me	H	-	-	90 ± 2*	42 ± 5*	50 ± 6
22	Me	H	H	Et	H	89 ± 9	9 ± 1	-	-	11 ± 3
23	Me	H	Et	H	H	39 ± 7	10 ± 2	-	-	15 ± 4
24	Et	H	H	Et	H	114 ± 18	3 ± 0	-	-	16 ± 2
25	Et	H	Et	H	H	64 ± 7	20 ± 5	-	-	29 ± 4
26	Et	H	Et	Et	H	-	-	69 ± 8	48 ± 15	42 ± 20
27	Et	H	H	Pr	H	69 ± 9	16 ± 5	-	-	32 ± 10

^aValues with standard errors (SEM) represent the mean value of at least three separate experiments with triplicate determinations and values without standard deviation represent a single experiment and values with * represent the mean value of two experiments with standard deviation. ^bAgonist efficacies were compared to that of dihydrotestosterone (100%) and antagonist efficacies were determined as a function (%) of maximal inhibition of dihydrotestosterone (EC₅₀). ^cAll EC₅₀ and IC₅₀ values were determined from full dose-response curves ranging from 10⁻¹² to 10⁻⁵ M in CV-1 cell. ^dA hyphen indicates an efficacy < 20% or a potency > 10000 nM. ^eThe radioligand used in the competitive binding assay was dihydrotestosterone.

References and Notes

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6. Preparation of **24** and **25**: A mixture of aniline (3.0 g, 32 mmol) and 2-pentenoic acid (2.5 g, 25 mmol) in toluene (20 mL) was heated at reflux for 20 h and removal of solvent followed by chromatography (EtOAc/hexane, 9/1) afforded 3.3 g (72%) of 3-phenylaminopentanoic acid. The acid was treated with PPA (20 mL) at 110 °C for 6 h and the reaction mixture was poured into ice water (50 mL) and then neutralized with Na₂CO₃ to pH 7. Extraction of the mixture with EtOAc (3 × 60 mL) followed by chromatography (EtOAc/hexane, 4/6) afforded 1,2,3,4-tetrahydro-2-ethyl-4-quinolinone (1.7 g, 54%) as a yellow solid. The quinolinone (1.0 g, 6.2 mmol) in THF (25 mL) was treated with di-*t*-butyl dicarbonate (1.6 g, 7.4 mmol) and DMAP (0.94 g, 7.4 mmol) at rt. for 6 h. Removal of solvent and chromatography afforded **4** (R¹ = Et, 1.6 g, 80%). To a solution of **4** (R¹ = Et, 0.20 g, 0.72 mmol) and iodoethane (0.50 mL, 6.3 mmol) in DMF (10 mL) was added NaH (60 % in mineral oil, 40 mg, 1.0 mmol) and the resulting mixture was stirred at rt. for 15 h. The reaction was quenched with water (10 mL) and extracted with EtOAc (2 × 25 mL). Removal of solvent and chromatography (EtOAc/hexane, 1/9) afforded a mixture of products, which was treated with TFA (1.0 mL) in methylene chloride (2.0 mL) for 3 h. The reaction was adjusted to pH 10 by 5% NaOH and extracted with EtOAc (2 × 20 mL). Chromatography (5–20% EtOAc/hexane, gradient) afforded a cis/trans mixture of 2,3-diethyl-1,2-dihydro-4-quinolones (85 mg, 58%) as a colorless oil. The quinolinone mixture (85 mg, 0.42 mmol) was treated with Et₃SiH (1.0 mL) and BF₃·OEt₂ (0.05 mL, 0.4 mmol) in CH₂Cl₂ (1.0 mL) at 100 °C for 15 h in a sealed tube followed by chromatography (EtOAc/hexane, 1/9) afforded a mixture of **5** (R¹ = R³/R⁴ = Et, 70 mg, 88%). The quinoline mixture (70 mg, 0.37 mmol) was converted to the final compounds according to a general nitration-hydrogenation-Knorr procedure⁴ in 30% (81%, 83%, 44%) three-step yield as a yellow solid (36 mg, 0.11 mmol) and HPLC separation provided the pure isomers. Compound **24**: mp, 263–264 °C; ¹H NMR (400 MHz, acetone-*d*₆) 10.71 (brs, 1 H), 7.30 (s, 1 H), 6.52 (s, 1 H), 6.41 (s, 1 H), 6.25 (brs, 1 H), 3.15 (m, 1 H), 2.95 (dd, *J* = 15.8, 4.8, 1 H), 2.58 (dd, *J* = 15.8, 6.2, 1 H), 1.71 (m, 1 H), 1.63 (m, 2 H), 1.49 (m, 1 H), 1.23 (m, 1 H), 0.99 (t, *J* = 7.3, 3 H), 0.97 (t, *J* = 7.3, 3 H); ¹³C NMR (100 MHz, acetone-*d*₆) 161.8, 149.0, 141.8, 125.5, 125.2, 122.8, 117.9, 114.3, 105.0, 96.8, 56.8, 56.6, 36.2, 28.7, 25.6, 11.5, 9.8. Compound **25**: ¹H NMR (400 MHz, acetone-*d*₆) 10.67 (bs, 1 H), 7.27 (s, 1 H), 6.52 (s, 1 H), 6.41 (s, 1 H), 6.30 (bs, 1 H), 3.35 (m, 1 H), 2.82 (m, 1 H), 2.71 (dd, *J* = 16.2, 8.1, 1 H), 1.89 (m, 1 H), 1.60–1.21 (m, 4 H), 1.01 (t, *J* = 7.5, 3 H) and 0.97 (t, *J* = 7.5, 3 H).
7. The cross-reactivity profile of **22** and **24** with other steroid receptors is consistent with related series; see ref. 3a and 9.
8. The substituent effects that switch agonist/antagonist activities have been seen in other steroid receptors, e.g. PR see (a) Cook, C. E.; Wani, M. C.; Lee, Y.-W.; Fail, P. A.; and Petrow, V. *Life Sciences* **1992**, *52*, 155, and ER see (b) Bowler J.; Lilley, T. J.; Pittam, J. D.; and Wakeling, A. E. *Steroids* **1989**, *54*(1), 71.
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